Volatile extraction from soybean plants infested with several herbivores

Extração de voláteis de plantas de soja infestadas por múltiplos herbívoros

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Abstract
The oral secretion of herbivores triggers chemical defenses in plants. When subjected to simultaneous or sequential attack by herbivores, plants emit volatile compounds of varying chemical nature, which can interfere with the attraction of natural enemies. This study investigates the profile of volatile compounds emitted by transgenic soybean (Bt M6210 PRO) plants infested with *Tetranychus urticae* Koch (Acari: Tetranychidae) and *Anticarsia gemmatalis* Hübner (Lepidoptera: Noctuidae). To this end, we used a method without leaf maceration. For plant infestation, 100 *T. urticae* mites per plant were used for one day and, subsequently, 2 *A. gemmatalis* caterpillars for 36 hours. Volatile compounds produced after infestation were analyzed by gas chromatography coupled with mass spectrometry. Principal component analysis showed differences between groups of compounds and explained 77.5% of the variation in volatiles. There are chemical differences between the emission of compounds according to the type of herbivory. It is concluded that soybean plants infested in a single (*T. urticae*) or multiple (*T. urticae* followed by *A. gemmatalis*) way trigger the production of volatile compounds that can be used as chemical traces to direct the foraging of natural enemies. The possible functions of volatiles produced after herbivory are discussed.

Additional keywords: *Anticarsia gemmatalis*; gas chromatography coupled with mass spectrometry; multiple attack; solid-phase microextraction; *Tetranychus urticae*.

Resumo
A secreção oral dos herbívoros aciona rotas químicas de defesas nas plantas. Ao serem submetidas ao ataque simultâneo ou sequencial de herbívoros, as plantas emitem compostos voláteis de variada natureza química que podem interferir na atração de inimigos naturais. Objetivou-se investigar o perfil dos compostos voláteis emitidos por plantas de soja transgênica (Bt) M6210IPRO infestadas com *Tetranychus urticae* Koch (Acari: Tetranychidae) e *Anticarsia gemmatalis* Hübner (Lepidoptera: Noctuidae) utilizando metodologia que não implicasse na maceração das folhas. Para infestação das plantas, foram utilizados 100 ácaros de *T. urticae* por planta durante um dia e, em sequência, 2 lagartas de *A. gemmatalis* durante 36 horas. Os compostos voláteis produzidos após infestação foram analisados por cromatografia gasosa acoplada à espectrometria de massas. A análise de componentes principais mostrou diferenças entre os grupos de compostos e explicou 77,5% da variação dos voláteis. Observou-se que existem diferenças químicas entre a emissão de compostos segundo o tipo de herbivoria. Conclui-se que plantas de soja infestadas por *T. urticae*, de maneira simples e múltipla (*T. urticae* seguido por *A. gemmatalis*), acionam a produção de compostos voláteis que podem ser utilizados como pistas químicas para direcionar o forrageamento de inimigos naturais. As possíveis funções dos voláteis produzidos após herbivoria são discutidas.

Palavras-chave adicionais: *Anticarsia gemmatalis*; cromatografia gasosa acoplada à espectrometria de massas; micro-extracção em fase sólida; múltiplo ataque; *Tetranychus urticae*.

Introduction
Plants have constitutive defenses against herbivores, which are characterized by the continued presence of morphological structures or chemicals that can negatively affect biological parameters related to herbivore development and reproduction (Coley & Barone, 1996). These structures and substances can be found in plants depending on the phenological stage (Coley & Barone, 1996). Plants can also respond to herbivory by triggering metabolic pathways for defense induction.
Materials and methods

Breeding of *Tetranychus urticae* and *Anticarsia gemmatalis*

Jack bean (*Canavalia ensiformis*) seeds were sown in plastic pots (6.3 L) containing substrate (Terral Solo®), which were kept in cages with anti-aphid screen in a greenhouse (25 ± 5 °C) for establishment of *T. urticae*.

A. *gemmatalis* caterpillars were bred by the team of the Biological Control Laboratory of Embrapa Maize and Sorghum, in Sete Lagoas city, Minas Gerais State. These caterpillars fed on artificial diet (Vilela et al., 2014) and sugary solution (50%) during the adult phase. Adult moths were placed in cylindrical PVC cages (30 cm in diameter and 20 cm in height) for copulation and oviposition. Napkins containing eggs were removed from the cages and stored in plastic bags at a controlled temperature (25 ± 2 °C). After hatching, the larvae were individualized in 50-mL lidded plastic containers containing artificial diet, remaining there until the pupa phase.

Sowing and maintenance of soybean plants

Plastic pots (1 L) with 500 grams of substrate Terral Solo® were used to sow 100 M6210 IPRO soybean plants expressing *Bacillus thuringiensis* (Bt) Cry1Ac protein. The plants were placed in cages with anti-aphid screen in a greenhouse. Irrigation and other cultural treatments were carried out whenever necessary. Soybean plants were used for the experiments when they reached the V3 development stage.

Plant infestation, collection and chemical analysis of volatiles

Infestations were performed simultaneously on soybean plants with 100 adult female *T. urticae* miles per plant for 24 hours and/or with two fourth-instar *A. gemmatalis* caterpillars for 36 hours prior to sampling. Headspace collection was performed according to the method proposed by Pereira et al. (2017) with adaptations. A plastic bag was used to pack the plants and an electric pump was used to inflate the air. The bag was completely heat-sealed and Teflon was used at the base to prevent air leakage. After exposure to herbivory, sterile syringes were used to collect volatiles in each treatment, which were transferred to headspace vials (25 mL), being identified and sealed (Figure 1).

Samples were analyzed via GC-MS system on a Trace GC Ultra gas chromatograph (Thermo Scientific, San Jose, CA) coupled to a Polaris Q mass spectrometer (ion trap) (Thermo Scientific, San Jose, CA), using solid-phase microextraction (SPME) in the headspace mode (Merkle et al., 2015). Polydimethylsiloxane/divinylbenzene (PDMS/DVB) semipolar fiber was used for vial exposure at 60 °C for 20 minutes.

Chromatographic conditions for the collection of soybean volatiles were: injector temperature = 200 °C; splitless injection; splitless time = 5 minutes; ion source temperature = 200 °C; interface temperature = 275 °C. The heating temperature of the equipment was 40 °C for 1 minute, followed by a gradient of 5 °C min⁻¹ to 110 °C, isothermal maintenance for 3 minutes, a new gradient of 7 °C min⁻¹ to 220 °C, with the temperature maintained for 1 minute and, finally, a gradient of 12 °C min⁻¹ to 245 °C.
temperature that was also maintained for 1 minute. The detector was kept in scan mode (fullscan, 30-300) using the electron impact (EI) ionization technique with 70 eV energy. The chromatographic column used was the HP-5 MS capillary column (5% phenyl - 95% methylpolysiloxane), containing the following dimensions: 30 m length, 0.25 mm internal diameter, and 0.25 μm film thickness (Agilent Technologies INC, Germany).

Compounds were identified by comparisons between mass spectra present in the NIST/EPA/NIH library (2005), based on retention time values obtained from the Xcalibur 1.4 program (Thermo Electron Corporation).

Figure 1 - A) Sealed soybean plants for the collection of volatiles. B) Collection of volatiles in a soybean plant.

Statistical Analysis

The results of the percentages of the relative areas of each sample were submitted to principal component analysis (PCA). The R program was used to perform statistical analysis and to elaborate the graphs (R Development Core Team, 2014).

Results

Eight compounds were identified and grouped into: (a) produced by clean plants infested with mites or with mites and caterpillars (camphene and 7-octen-3-ol, 2,6-dimethyl); b) produced under all conditions (2-octenal); c) produced only in caterpillar infestations with or without mites (lavandulol and cyclohexanol, 2-methyl-5-(1-methylethyl); d) produced only by clean plants (dodecene); e) produced only by clean plants or caterpillar-free plants infested with mites (geranyl isovalerate); f) produced only by caterpillar-infested plants without mites (linalool) (Table 1).

Table 1 - Volatile organic compounds (VOC) found in soybean plants (M6210IPRO): Cleaned soybean plants (S), infested with arid-brindle, *Tetranychus urticae* (SA), infested with two *Anticarsia gemmatalis* (SL) and *T. urticae + A. gemmatalis* (SAL) on the same plant.

<table>
<thead>
<tr>
<th>n</th>
<th>VOC</th>
<th>Class</th>
<th>Chemical formule</th>
<th>S</th>
<th>SA</th>
<th>SL</th>
<th>SAL</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>Camphene*</td>
<td>Monoterpenes</td>
<td>C10H16</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>2-Octenal*</td>
<td>Jasmonate*</td>
<td>C8H14O</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>3</td>
<td>Lavandulol</td>
<td>Monoterpenes</td>
<td>C10H22O</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>4</td>
<td>Dodecene*</td>
<td>Jasmonate*</td>
<td>C12H24</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>5</td>
<td>Cyclohexanol,2-metil-5-(1-methylethyl)</td>
<td>Monoterpenes</td>
<td>C10H20O</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>6</td>
<td>Geranyl isovalerate</td>
<td>Monoterpenes</td>
<td>C15H26O2</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>7</td>
<td>7-Octen-3-ol,2,6dimethyl</td>
<td>Jasmonate*</td>
<td>C10H20O</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
</tr>
<tr>
<td>8</td>
<td>Linalool*</td>
<td>Monoterpenes</td>
<td>C10H18O</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

Total of compounds 5 4 5 5

S = uninfested soybean; SA = soybean infested by the arid-brindle; SL = soybean infested by the soybean caterpillar; SAL = soybean infested by the arid-brindle and the soybean caterpillar. * Volatile organic compounds present in soybean plants previously described in the literature.

Principal component analysis of chemical parameters related to volatile composition in clean and infested soybean leaves explained 77.5% of the data variation. The samples of the three plants infested with *A. gemmatalis* and under multiple herbivory (*T. urticae* and *A. gemmatalis*) were grouped in different positions, indicating that there was a difference between the volatile compounds emitted in both situations, whereas the samples of clean plants approached those with *T. urticae* infestation (Figure 2).
Figure 2 - Principal components analysis of volatile compounds present in soybean plants without infestation (○), soybean infected with T. urticae (Δ); soybean infested with A. gemmatalis (●) and soybean infected with T. urticae + A. gemmatalis (▲) detected via GC/MS.

Figure 3 shows the contribution of compounds to group formation, where monoterpenes lavandulol and cyclohexanol are predominant in multiple infestations (mite + caterpillar). Caterpillar-infested soybean plants activate the jasmonate route with the production of dodecene, and the isoprene route with the formation of linalool.

Figure 3 - Analysis of major components of volatile compounds present in soybean plants detected via GC/MS. The numbers used for compounds identification are the same of those described in Table 1.
Discussion

Compounds camphene, 2-octenal, dodecene, and linalool, found in the present work, were also identified in soybean plants in several studies evaluating the emission of volatile compounds (Rosario et al., 1984; Liu et al., 1989; Damiani et al., 2000; Boue et al., 2003; Boer et al., 2004; Moraes et al., 2005; Zhu & Park, 2005; Rostas & Eggert, 2008; Michereff et al., 2011; Cai et al., 2015). Moreover, compounds 2-octenal, dodecene, and 2,6-dimethyl-7-octen-3-ol, derived from jasmonate, also strongly suggest indirect induced defense, as the jasmionic acid route has been described as one of the major chemical routes of volatiles (Heil, 2014).

In the volatile analysis of clean soybean plants, compound dodecene, found exclusively in cultivar PI 227687, repelled caterpillar Trichoplusia ni (Liu et al., 1989). In turn, soybean cultivars that did not release dodecene were attractive to T. ni. Compound dodecene is shown to be a signaling molecule for host localization by the herbivore. Infestation conditions cause the plant to interrupt the production of this compound to benefit the herbivore so that new herbivores do not colonize the same host.

The predatory mite Phytoseiulus persimilis preferred volatiles of bean plants with low density of S. exigua (2 caterpillars per plant), repelling plants with a high density of the insect (100 caterpillars per plant). However, predators did not prefer this treatment when caterpillar feces were not removed. In that study, the authors showed that caterpillar fecal volatiles mediate the preference. Such observation is pertinent because caterpillar feces remained on the soybean leaves during the experiment, thus fecal volatiles may also have been captured.

Compound linalool, found in soybean plants infested with A. gemmatalis, was also found in soybean plants infested with Spodoptera frugiperda (Rostas & Eggert, 2008). This compound may be key in indirect induced defense and assist in the foraging of natural enemies by plants infested with prey herbivores. Compound linalool was not produced by caterpillar-infested plants that were previously exposed to mite herbivory. It is suggested that some mechanism of volatile emission regulation may occur by mite herbivory.

Herbivory is shown to alter the chemical profile of volatiles emitted in soybean plants, and these volatiles possibly characterize the triggering of indirect induced defense.

Conclusions

The developed method allows to affirm that the compounds found are released by soybean plants after herbivore manipulation, without possible interference of mechanical damage caused in the maceration process. Future work should be developed testing the preference for natural enemies to clarify field foraging.

References


